

Triple antibiotic therapy with ceftolozane/tazobactam, colistin and rifampin for pan-resistant *Pseudomonas aeruginosa* ventilator-associated pneumonia

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ABSTRACT

Emergence of multi-drug resistant microorganisms, such as pan-resistant *Pseudomonas aeruginosa*, has recently created a therapeutic challenge in ICU patients worldwide. New antipseudomonal antibiotics, like ceftolozane/tazobactam, have been developed to meet this challenge. This drug does not demonstrate cross-resistance with other antimicrobial classes, like carbapenems, because of its enhanced binding affinity to the penicillin-binding proteins. A Phase III, multicenter, prospective, randomized, double blind study has been initiated to evaluate the safety and efficacy of ceftolozane/tazobactam in ventilator-associated pneumonia (VAP). We present a case of VAP due to pan-resistant *Pseudomonas aeruginosa* in a patient with advanced multiple sclerosis. He was treated with ceftolozane/tazobactam in combination with colistin and rifampin for synergistic effect. Within two weeks of treatment, he had significant improvement in his leukocytosis and chest infiltrates, and his ventilator settings were adjusted to their baseline settings. This case illustrates the importance of using this novel antipseudomonal antibiotic to treat bacteria that are resistant to a wide spectrum of antibiotics, including carbapenems. Other antibiotics, like colistin and rifampin, can be used for synergism until more data are collected from trials evaluating the efficacy of monotherapy with this novel antibiotic for VAP.

Key words: pan-resistant *Pseudomonas*, novel antipseudomonal antibiotic, combination therapy

INTRODUCTION

Pseudomonas aeruginosa is responsible for a significant percentage of nosocomial infections, including pneumonia, bacteremia, and urinary tract infections. The ability of a single strain of *P. aeruginosa*

to acquire antimicrobial resistance via multiple mechanisms has made *P. aeruginosa* especially difficult to treat.¹ Ceftolozane/tazobactam is a novel antibacterial agent and β -lactamase-inhibitor combination that has appreciable activity against wild-type *Enterobacteriaceae* and *P. aeruginosa*.² Combination therapies based on polymyxin E (colistin), rifampin, and carbapenems have been used in many cases to effectively treat multidrug resistant Gram negative infections.³

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CASE

A 69-year-old man had progressive advanced multiple sclerosis. His respiratory system had been compromised by his disease, and he had undergone permanent tracheostomy. During the past year he became a resident of a nursing home and required assist-control mode ventilation with a backup rate of 12, a tidal volume of 600 mL, a positive end-expiratory pressure (PEEP) of 5 cmH₂O, and a FiO₂ of 40%. He became tachypneic and somnolent and developed increased tracheostomy secretions. He was admitted to the medical intensive care unit; his initial work up revealed a WBC of 29.9 k/μL and urine WBC of 104/HPF. His chest x-ray showed bilateral interstitial infiltrates with a right lower lung field predominance (Figure 1). Empiric therapy with vancomycin and piperacillin/tazobactam was started. The patient required a backup rate of 20 and a PEEP of 10 to maintain oxygen saturation above 90 percent. On hospital day

three the final urine culture grew Extended Spectrum Beta-Lactamase-producing (ESBL) *Proteus mirabilis* and *Providencia stuartii* (Figure 2). A culture of tracheostomy aspirate began growing Gram negative bacilli that were oxidase positive and non-lactose fermenting. Piperacillin/tazobactam was changed to meropenem, and vancomycin was stopped. By day five his WBC had dropped to 13 k/μL, but he still required the same back up rate and PEEP on mechanical ventilation. His final tracheobronchial culture showed pan-resistant *Pseudomonas aeruginosa* that was also resistant to meropenem (Figure 3). The patient was started on ceftolozane/tazobactam 1000/500 mg IV every 8 hours, colistin 150 mg IV every 12 hours, and rifampin 300 mg every 12 hours via PEG tube. Within two weeks, his WBC decreased to 7k/μL, the back-up rate was decreased to 12, PEEP was decreased to 5 cmH₂O, and the chest x-ray showed near complete resolution of infiltrates (Figure 4).

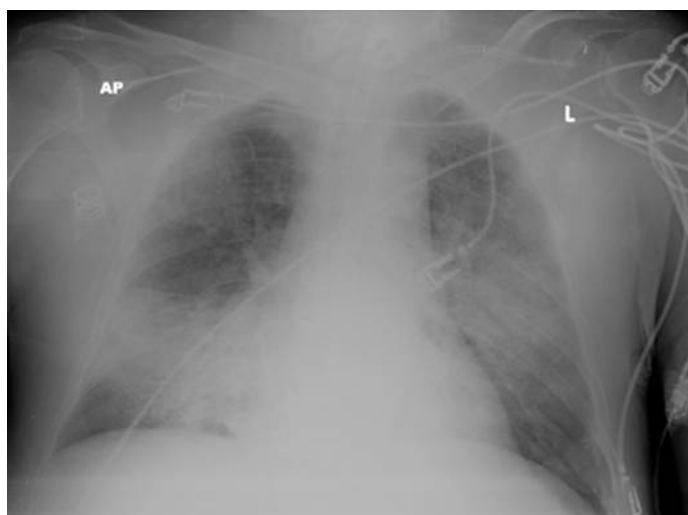


Figure1: Chest x-ray on admission showing bilateral interstitial infiltrates, especially in the right lower lung field.

CULTURE URINE Final		PROTEUS MIRABILIS		PROVIDENCIA STUARTII		
Organism 1	COLONY COUNT:	25,000-50,000 CFU/ML		50,000-75,000 CFU/ML		
Organism 2	COLONY COUNT:					
(PROTEUS MIRABILIS) IS (EXT BETA LACTAMASE POSITIVE)						
	P MIRABIL	M.I.C.	BX	P STUARTII	M.I.C.	BX
AMPICILLIN	>16	R	>16	R	>16	R
AZTREONAM	8	R	<=4	S	<=4	S
CEFAZOLIN	>16	R	>16	R	>16	R
CEFEPIME	>16	R	<=4	S	<=4	S
CEFOTAXIME	>32	ESBL	8	S	8	S
CEFOXITIN	<=8	S	<=8	S	<=8	S
CEFTAZIDIME	4	ESBL	8	S	8	S
CEFTRIAXONE	>32	R	<=8	S	<=8	S
CEFUROXIME	>16	R	>16	R	>16	R
CIPROFLOXA	>2	R	>2	R	>2	R
DORIPENEM	<=0.5	S	<=0.5	S	<=0.5	S
ERTAPENEM	<=1	S	<=1	S	<=1	S
GENTAMICIN	>8	R	<=4	S	<=4	S
LEVOFLOXACIN	>4	R	>4	R	>4	R
MEROPENEM	<=1	S	<=1	S	<=1	S
NITROFURAN	>64	R	>64	R	>64	R
PIP/TAZO	<=16	S	<=16	S	<=16	S
TETRACYCLI	>8	R	>8	R	>8	R
TIGECYCLINE-E	>8	R	2	S	2	S
TOBRAMYCIN	>8	R	>8	R	>8	R
TRIM/SULFA	>2/38	R	<=2/38	S	<=2/38	S

Figure2: Urine culture showing *Proteus mirabilis* (ESBL) and *Providencia stuartii*.

RESPIRATORY CULTURE Final
 Organism 1 PSEUDOMONAS AERUGINOSA
 COLONY COUNT: MODERATE
 SENSITIVITY TO FOLLOW

FEW USUAL RESPIRATORY FLORA.

PSEUDOMONAS AERUGINOSA IS MULTI-DRUG RESISTANT ORGANISM.

{PSEUDOMONAS AERUGINOSA} IS {MULTI-DRUG RESISTANT ORGANISM}
 COLISTIN: 14 MM -SUSCEPTIBLE
 TIGECYCLINE MIC: 256
 ZERBAXA MIC: 1.5

1. PSEUDOMONAS AERUGINOSA	RX	AB	Cost	M.I.C.
AZTREONAM	I	A	16	
CEFEPIME	R	A	>16	
CEFOTAXIME	R	A	>32	
CEFTAZIDIME	R	A	>16	
CEFTRIAKONE	R	A	>32	
CIPROFLOXA	R	A	>2	
GENTAMICIN	R	A	>8	
LEVOFLOXACIN	R	A	>4	
MEROPENEM	R	A	>8	
PIP-TAZO	R	A	>64	
TOBRAMYCIN	R	A	>8	
TRIM/SULFA	R	A	>2/38	

Figure3: Tracheobronchial aspirate culture showing pan-resistant *Pseudomonas aeruginosa*.

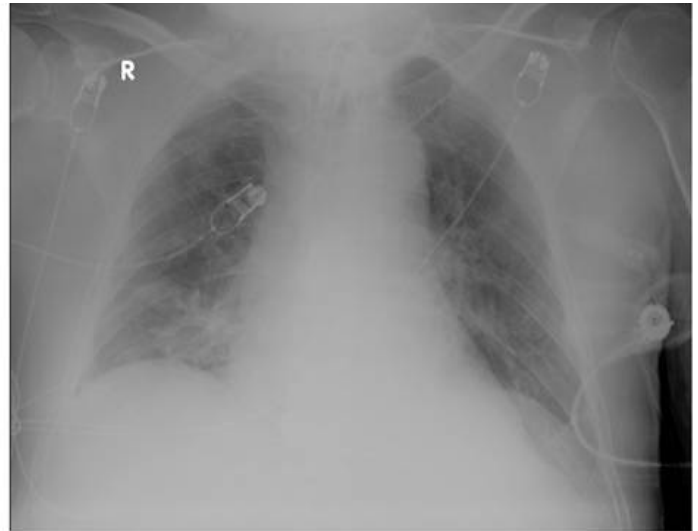


Figure4: Repeat chest x-ray after two weeks therapy with the triple antibiotics showed near complete resolution of infiltrates.

DISCUSSION

Despite the wide distribution of *P. aeruginosa* in the environment, this microorganism rarely colonizes humans. However, the incidence of colonization increases significantly in hospitalized patients. In some hospitals, *P. aeruginosa* is the initial causative organism of infection, particularly in urinary and lower respiratory tract infections. Resistance of *P. aeruginosa* to antipseudomonal drugs has increased throughout the world. The mechanisms of resistance are complex and multifactorial and involve acquisition of genes (mainly against beta-lactams and aminoglycosides), chromosomal gene mutations (target site mutations affecting fluoroquinolones), and up-regulation of multidrug efflux pumps. Carbapenems have been one of the most important classes of antibiotics used in the empiric treatment of nosocomial infections, especially in severe cases. However, resistance of *P. aeruginosa* to imipenem and meropenem is increasing. The main mechanism of resistance to carbapenems in the United States is the loss of the OprD outer membrane porin protein. In other countries production of metallo-beta-lactamases also has a major role.^{4,5}

Ceftolozane/tazobactam (formerly CXA-201)

is a novel antibacterial and β -lactamase-inhibitor combination with the potential to meet the challenges created by multidrug resistant strains of *P. aeruginosa*. It has demonstrated good stability to AmpC β -lactamases and is less affected by changes in porin permeability and efflux pumps as a result of its strong binding of select penicillin-binding proteins (PBPs).⁶ More specifically, in comparison to ceftazidime and imipenem, ceftolozane has greater affinity for all essential PBPs (1b, 1c, 2, and 3).⁷

Currently, a trial is enrolling subjects in a prospective, randomized, double blind, multicenter, phase 3 clinical study to assess the safety and efficacy of IV ceftolozane/tazobactam compared to IV meropenem in adult patients with ventilated nosocomial pneumonia. Subjects receive either ceftolozane/tazobactam 3 g (administered IV over 60 minutes) every 8 hours for 8 days (14 days for *Pseudomonas aeruginosa*) or meropenem 1 g (administered IV over 60 minutes) every 8 hours for 8 days (14 days for *P. aeruginosa*).⁸ In part, the rationale for employing higher doses of ceftolozane stems from the results of an experimental rabbit pneumonia model that showed significantly

greater reduction in pulmonary bacterial load with ceftolozane human-equivalent doses of 2000 mg IV every 8 hours compared to ceftolozane human-equivalent doses of 1000 mg IV every 8 hours.⁹ Ceftolozane/tazobactam 1.5 g administered every 8 h via a 60 minute infusion was used in our patient as acceptable plasma and epithelial lining fluid concentrations were achieved in a previous study with this dose.¹⁰ Our literature review found several studies citing the synergistic activity of dual combination therapy regimens consisting of colistin plus rifampin, colistin plus carbapenem, and rifampin plus carbapenem in the treatment of multidrug resistant Gram negative bacteria, but only one case report using triple combination therapy (colistin, rifampin and meropenem) has been described in the treatment of multidrug resistant *A. baumannii* infection.³

Our patient was already being treated with meropenem for his ESBL urinary tract infection, and his *Pseudomonas* isolate was resistant to the carbapenem class. The *Pseudomonas* sensitivity to colistin was evaluated by the disc susceptibility testing method, which showed a zone of inhibition of 14 mm. Interpretative criteria for disc susceptibility testing of colistin are not available from the Clinical and Laboratory Standards Institute (CLSI) and interpretations of inhibition zone size are based on product literature. A previous study showed that disc diffusion remains an inherently unreliable susceptibility testing method and failed to detect colistin resistance compared to dilution-based methods, particularly for *P. aeruginosa*.¹¹ We elected to use ceftolozane/tazobactam as the backbone of our antibiotic treatment for this case of pan-resistant *Pseudomonas* VAP since the isolate was sensitive to this antibiotic with a MIC of 1.5 (with a MIC cutoff limit of 4 mg/L considered to be resistant by the E test strip dilution method) and that information was available within 5 days of sample collection. We added colistin and rifampin for synergistic effect. His creatinine was closely monitored, as nephrotoxicity has been reported in some cases with the use of intravenous colistin. Rifampin, when combined with colistin, has been shown to have *in vitro* and *in vivo* synergistic effects in the treatment of multidrug resistant Gram negative bacteria.¹² In our patient, this triple

combination therapy was effective and well tolerated.

CONCLUSIONS

Until controlled trials evaluating the effectiveness of monotherapy with novel antipseudomonal antibiotics like ceftolozane/tazobactam in the treatment of multidrug resistant Gram negative VAP are available, triple antibiotic therapy with the addition of colistin and rifampin for synergy is a promising option on a case by case basis.

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